

QUESTION 9: Is there evidence that interference with bacterial communication by blocking quorum sensing molecules can minimize biofilm formation *in vivo*?

Authors: Alex C McLaren, Garth D Ehrlich

Response:

In vivo animal studies have demonstrated that interference with quorum sensing signals/molecules in some infections leads to decreased biofilm formation. There are contradictory results in *Staphylococcus* species. However, there are no clinical studies demonstrating this phenomenon.

Level of Evidence: Limited

Delegate Vote: Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

Post Meeting Rationale:

In Feb 2018 NCBI, EMBASE and SCOPUS databases were searched for English language publications during the previous 5 years in which the abstract contained the following terms: “biofilm” and “quorum” and “*in vivo*”. These papers were reviewed to identify data specifically related to interference of quorum sensing the caused alteration in biofilm formation. All *in vitro* basic science studies were excluded and there were no clinical studies. There were 7 *in vivo* investigations reported during the last five years¹⁻⁷ (Table 1). In addition, there have been reports of quorum sensing inhibitor and quorum quenching studies presented at scientific meetings utilizing multiple *in vivo* models⁸.

The experimental strategy varies. *In vitro* data are relied upon to identify the molecular mechanism leading to interference with quorum sensing that causes decreased biofilm formation, whether it be blocking the signaling peptide production, blocking receptors, or active initiation an antagonist signals by the agent. *In vivo* studies are then done to confirm that the agent decreases biofilm formation.

While there is extensive *in vitro* and *in silico* work being done and reported on quorum sensing and anti-quorum sensing molecules, otherwise known as quorum quenching, there are limited *in vivo* data and none of the anti-quorum sensing strategies are ready for widespread clinical application.

Table 1: In vivo investigation during the last 5 years

Study	Animal Model	Agent	Mechanism	Clinical effect
1	Medaka fish peritoneal catheter infection	3-Phenyllactic Acid (PLA)	Antagonistically binds to quorum sensing receptors RhIR and RqsR, blocking initial attachment of <i>Pseudomonas aeruginosa</i> (PA01) thereby delaying biofilm formation	Decreased biofilm formation
2	Mouse gingivitis	Quorum Sensing Inhibitors (furane compounds, d-ribose)	Interfere with AutoInducer-2	decreased colony counts and alveolar bone loss
3	Round worm survival (<i>Caenorhabditis elegans</i>)	Pyrrolo[1,2-a]pyrazine-1,4-dione, hexahydro-3-(2-methylpropyl) from <i>Alcaligenes faecalis</i>	Modulate expression of quorum sensing (QS) regulators luxT and lafK	Increased survival from <i>V. alginolyticus</i> infection
4	Round worm survival (<i>Caenorhabditis elegans</i>)	Sub-inhibitory concentration of ceftazidime	inhibition of QS regulated virulence traits and biofilm formation; binds to the las and pqs QS receptors in <i>P. aeruginosa</i>	Increased survival
5		Acylase	Degrades Quorum sensing peptides	Delay biofilm formation for <i>S. aureus</i> and <i>P. aeruginosa</i> for up to 7 days
6	larval oyster mortality	<i>Phaeobacter gallaeciensis</i> S4Sm	Down regulate pathogen virulence genes	Decreased mortality from <i>V. tubiashii</i> infection
7	Wistar rat pyelonephritis	Phytol	down regulate <i>offimA</i> , <i>fimC</i> , <i>flhC</i> , <i>flhD</i> , <i>bsmB</i> , <i>pigP</i> <i>shlA</i> genes in <i>S. Marcescens</i> leading to decreased biofilm formation and virulence factor production	Decreased bacterial counts and virulence enzymes (lipase and protease) decreased inflammatory markers (MDA, NO, MPO) and histologically no acute inflammation

References:

1. Chatterjee M, D'Morris S, Paul V, et al. 2017. Mechanistic understanding of Phenyllactic acid mediated inhibition of quorum sensing and biofilm development in *Pseudomonas aeruginosa*. *Appl. Microbiol. Biotechnol.* 101(22):8223–8236.
2. Cho Y-J, Song HY, Ben Amara H, et al. 2016. In Vivo Inhibition of *Porphyromonas gingivalis* Growth and Prevention of Periodontitis With Quorum-Sensing Inhibitors. *J. Periodontol.* 87(9):1075–1082.
3. Durai S, Vigneshwari L, Balamurugan K. 2013. *Caenorhabditis elegans*-based in vivo screening of bioactives from marine sponge-associated bacteria against *Vibrio alginolyticus*. *J. Appl. Microbiol.* 115(6):1329–1342.
4. Husain FM, Ahmad I, Baig MH, et al. 2016. Broad-spectrum inhibition of AHL-regulated virulence factors and biofilms by sub-inhibitory concentrations of ceftazidime. *RSC Advances* 6(33):27952–27962.
5. Ivanova K, Fernandes MM, Francesko A, et al. 2015. Quorum-Quenching and Matrix-Degrading Enzymes in Multilayer Coatings Synergistically Prevent Bacterial Biofilm Formation on Urinary Catheters. *ACS Appl Mater Interfaces* 7(49):27066–27077.
6. Rowley D., Zhao W., Yuan T., et al. 2015. Mechanisms of microbe-microbe-host interactions in a probiont-pathogen-bivalve model. *Planta Med.* 81(11) Available from: <http://www.embase.com/search/results?subaction=viewrecord&from=export&id=L71958553>.
7. Srinivasan R, Mohankumar R, Kannappan A, et al. 2017. Exploring the Anti-quorum Sensing and Antibiofilm Efficacy of Phytol against *Serratia marcescens* Associated Acute Pyelonephritis Infection in Wistar Rats. *Front Cell Infect Microbiol* 7:498.
8. Coenye T. 2013. Quorum quenching and quorum sensing inhibition to fight biofilm-related infections. *Int. J. Antimicrob. Agents* 42((Coenye T., Tom.Coenye@UGent.be) Laboratory of Pharmaceutical Microbiology, Ghent University, Belgium):S15–S16.